



E-ISSN: 2709-944X
P-ISSN: 2709-9431
JRM 2022; 3(2): 91-95
© 2022 JAMR
www.microbiojournal.com
Received: 03-08-2022
Accepted: 05-09-2022

Baydaa Ghanim Mohammed
Department of Biology,
College of Education for girls,
University of Mosul, Iraq

Jassim Fatehi Ali
Department of Biology,
College of Education for Pure
Science, University of Mosul,
Iraq

Ibtisam Kareem Mohaisen
Department of Basic of
Medical Sciences, College of
Dentistry, University of Misan

Identification and antibiogram of aerobic and anaerobic bacteria isolated from frozen fish retailed at public markets in Misan city, Iraq

Baydaa Ghanim Mohammed, Jassim Fatehi Ali and Ibtisam Kareem Mohaisen

DOI: <https://doi.org/10.22271/micro.2022.v3.i2b.54>

Abstract

The aim of the current study was to identify the bacteria that can be isolated from various samples of frozen fish and to examine their antibiograms. 45 samples of three types of frozen imported fish (Zuri, filli fish, *Pangasius hypophthalmus*) (15 of each sample) were collected from different markets in Misan Governorate, during the periods from 1/12/2020 to 15/1/2021. The isolation and identification of bacteria as well as bacterial sensitivity test to antibiotics were performed. The results of this study revealed the presence of different bacterial types in frozen fish samples, the types of the detected bacteria involved the following kinds: Coagulase-negative staphylococcus (2 isolates), Enterobacter aerogenes (1 isolate), and E. coli (1 isolate), but *Leuconostoc citreum* was not isolated. The amount of the isolation were different in amount depending on the types of fish studied. The presence of the detected bacteria was more prevalent in zuri than in white fish fillet and *Pangasius hypophthalmus* fish. The prevalence of both E.coli bacteria and Enterobacter aerogenes was the highest in zuri, while the contamination with Enterobacter aerogenes, Coagulase negative staphylococcus and *Leuconostoc citreum* were the lowest in both *Pangasius hypophthalmus* fish and white fish fillet. In conclusion, different types of bacteria that showed various degree of resistance to antibiotics were isolated from the three types of studied fish.

Keywords: Isolation contaminated bacteria, frozen fish, Food Safety, Antibiotic Sensitivity

1. Introduction

Fish is considered as an important food which has high nutritive value, as it is a source of low price animal protein as well as other important nutrients like calcium, phosphorus, and vitamins (Balami *et al.*, 2019) [1]. The aquatic environment and delicate tissues of fish are extremely vulnerable to be invaded by microbes (Begum *et al.*, 2021) [5]. During the past three decades, the eating habits of human were changed considerably, especially in East Asia particularly in China and East/North Africa region. Recently and according to a survey, 20 kgs of fish are consumed by the average person per year, which represents about 6.7% of all the protein that is ingested by people worldwide (FAO, 2016) [6]. Due to perishable nature of fish, its flavor and aroma are mainly affected by the temperature of preservation following being caught (Shikha *et al.*, 2022) [2]. Approximately, bacterial counts of 10^7 cfu/g of the fish lead to the production of an off flavor and odor (Gelman *et al.*, 2001) [3]. The majority of the transmitted diseases from fish to humans through zoonotic agents are bacterial, and infection is contracted when diseased fish or fomites are handled and result in cuts, abrasions and even puncture wounds to the skin (Boylan, 2011) [7]. Consumption of raw fish or fish with little to no processing results in bacterial contamination which either caused by direct contamination of fish by water or it may cause by secondary contamination during handling, processing, storage, distribution and preparation (Yemmen and Gargouri, 2022) [10]. Some bacterial species are able to cause disease in both people and fish. Without any clear signs of the disease, they may be separated from fish. There is a link between the following bacteria and human illnesses and poisonings: *Aeromonas* species, *Salmonella* species, *Staphylococcus aureus*, *Clostridium botulinum*, *C. perfringens*, *Campylobacter jejuni*, *Delftia acidovorans*, *Edwardsiella tarda*, *Legionella pneumophila*, and *Plesiomonas shigellois*'s are among the bacteria that have been identified. *Streptococcus ini ae*, *Photobacterium damsela*, VI (Novotny *et al.*, 2017) [9]. *Pseudomonas*, *Aeromonas*, and *Enterobacter* can be considered the most common bacterial species that are responsible for fish rotting (Anagnostopoulos *et al.*, 2022) [4].

Correspondence

Baydaa Ghanim Mohammed
Department of Biology,
College of Education for girls,
University of Mosul, Iraq

Based on the outcomes of various research, fish have bacterial colonies on their skin, gills, digestive tract and light-emitting organs (Zaccone *et al.*, 2022) [8]. The direct transmission of fish-borne zoonotic illnesses results from consumption of raw or inadequately cooked fish meat. Indirect transmission is caused by presence of the contaminated water in the vicinity of diseased fish (Lehel *et al.*, 2021) [11]. It is challenging mission to accomplish this goal due to a scarcity of information related to the microbiological quality of fish. There is trouble of processing frozen fish for export because of adhering to the HACCP protocol requirements. The goal of the current study was to ascertain the microbiological quality of fish following cooking and drying. Additionally, the study examined the isolated bacteria's unique pattern of antibiotic susceptibility. The current study was also conducted to isolate and detect bacterial infections from frozen fish samples and to raise awareness among local consumers and recipients.

2. Materials and Methods

Collection of fish samples

A total of 45 random samples of frozen fish of Zuri, filli fish and *Pangasius hypophthalmus* (15 of each) were collected from different fish markets at Misan Governorate. Each sample was kept individually in separated plastic bag and transferred directly to the laboratory in an insulated box under complete aseptic condition without any delay. The collected samples were examined bacteriologically as soon as possible.

Bacterial diagnosis

Official methods (AOAC) were adopted to detect bacterial contamination. Following bringing of samples to the laboratory, they were collected at a weight of 20 grams of each fish and from different parts of the body and were placed in the center of the liquid media (nutrients broth). Other 20 grams samples were placed in the middle of a liquid media which contained 5 ml of nutrient broth, then the decimal dilution was made for each sample of fish in nutrient broth 10^{-2} , 10^{-3} , 10^{-4} the samples were later cultured in the nutrient broth agar and were incubated at of 37 °C m for one day for salmonella isolates were cultured on the special media.

Isolation and Identification of Bacteria

The morphology of bacterial colony was evaluated by assessment of colony size, shape, density, and color. Some biochemical tests were also used for isolation and identification of bacteria such as sugar test, Gram staining method, motility and indole test.

The precise species of bacteria were determined by subculture of the suspected colony in nutrient agar and nutrient broth media. Finally, the pure cultivation was performed by using a zigzag slant. The different biochemical tests such as gram staining and MR-VP tests, were accomplished in a completely sterile environment. Sensitivity test was used to determine sensitivity of the bacterial to antibiotics. Streaking on different solid agars was performed under laminar air flow.

Identification of isolates

According to manufacturer guidelines, the identification of all fish-isolated bacteria was achieved by using the Vitek equipment (bio Merieux Vitek Systems Inc., Hazelwood, Mo.).

Chemical analysis

The procedures used for calculation of the amounts of protein, fat, moisture and ash were recommended by the American Organization for Chemical Analysis (Association of Official Agricultural Chemists).

Statistical analysis

The average and standard deviation for the total number of aerobic plates were obtained. The Social Package of Social Sciences (SPSS) versiona22 was used to examine the data, and independent samples T-test was used to calculate statistical differences (Al-Rawi and Khalf Allahs, 2000).

3. Results and discussion

The tests results of microbial loads, staining, culture, biochemistry, antibiotic sensitivity pattern, and percentage of the isolated bacterial incidence were shown in distinct tables and discussed below under the following areas. The findings of this study revealed the presence of different bacterial kinds in frozen fish samples that were shown in Table 1, the types of the detected bacteria were as the following: Coagulase-negative staphylococcus (2 isolates), Enterobacter aerogenes (1 isolate), and E. coli (1 isolate), but Leuconostoc citreum was not detected. These isolations were in different amounts relying on the kinds of fish studied, the presence of the detected bacteria was more prevalent in zuri than in white fish fillet and *Pangasius hypophthalmus* fish. It was found that the presence of both E.coli bacteria and Enterobacter aerogenes was the highest in zuri, while the contamination with Enterobacter aerogenes, Coagulase negative staphylococcus and Leuconostoc citreum were the lowest in both *Pangasius hypophthalmus* fish and white fish fillet. These findings were similar to the study conducted by (Hussein *et al.*, 2009) [24].

Table 1: Percentages of bacterial types isolated from samples of Sfrozen fish

Bacterial types	Total number of bacterial in frozen <i>Clupeonella cultriventris</i> (zuri) fish	Number of isolatets	Bacterial count	Total number of bacterial isolates in frozen <i>Pangasius hypophthalmus</i> fish	Bacterial count	Total number of bacterial isolates in frozen white fish fillet	Bacterial count
E.coli	14	1	2x10 ⁵	6	2.5x10 ⁵	7	3x10 ⁶
Enterobacter aerogenes	10	1	1x10 ⁶	2	3x10 ⁵	7	2x10 ⁵
Coagulase negative staphylococcus	7	2	3x10 ⁴	2	2x10 ⁶	3	3x10 ⁴
Citrobacter freundii	5	1	4x10 ⁵	2	2x10 ⁷	2	2x10 ⁵
Leuconostoc citratum	7	0		2	2x10 ⁶	5	3x10 ⁵

The same table showed that the rates of several microbe species in white fish fillet and *Clupeonella cultriventris* (zuri) fish. It was observed that the prevalence of *E. coli* and *Leuconostoc citreum* bacteria had increased which could be attributed to the very high temperatures in different central markets. These markets seem not to be adhered to the practice of storing and marketing fish by following particular cooling methods. Finally, these circumstances promote the growth of bacteria by contributing to the melting and warming of the outer surface, which allow the growth of a several germs and produce perishable and unfit materials for human use (hameed Albassamet *et al.*, 2021) [26]. The prevalence of variety coliforms, heterotrophic indicator microorganisms and pathogenic strains in the ice which is used to freeze fish and the other seafood has been studied by Falcao *et al.* (2002) [25], who suggests that the ice used to refrigerate seafood could be a source of coliform bacteria that leads to human infection. As a result, some of the contamination detected in the current investigation could be related to the ice that is used for freeze during processing. Numerous enteric bacteria were detected, which indicated

the level of handler cross contamination. Their presence is of possible risk to people, especially those suffering from weakened immune systems, such as those with HIV/AIDS. To guarantee the safety of fish consumption, strict rules must be followed which are related to the registration of fishermen, traders, education and the necessity of monitoring sanitation at trading locations and using ice properly (Mhango *et al.*, 2010) [29]. In this study, the results related to the test of antibiotic resistance in bacterial isolated in *Pangasius hypophthalmus* revealed that that most of the *E. coli* isolates were resistant to Ciprofloxacin (100%) as shown in Table 2. The resistance to Oxacillin (40%) and Erythromycin (40%) was found to be lower. Enterobacter aerogenes showed resistance to Gentamycin (85.7%), Erythromycin (85.7%), Ampicillin (71.5%) and Ciprofloxacin (70%). Whereas the resistance to Oxacillin (57.1%), Tetracycline (57.1%) and Ciprofloxacin (50%) was lower. Coagulase negative staphylococcus, *Citrobacter freundii* and *Leuconostoc citreum* were lower resistant to all antibiotics.

Table 2: Percentages of antibiotic resistance in bacterial isolated in *Pangasius hypophthalmus*.

Bacterial type	Ampicillin	Oxacillin	Gentamycin	Erythromycin	Ciprofloxacin	Tetracycline	Ciprofloxacin
<i>E. coli</i>	0	40	0	40	100	0	0
Enterobacter aerogenes	71.5	57.1	85.7	85.7	70	57.1	50
Coagulase negative staphylococcus	50	50	0	50	50	0	50
<i>Citrobacter freundii</i>	0	0	0	0	30	0	25
<i>Leuconostoc citreum</i>	34	30	40	0	25	0	25

There were various gram-positive and gram-negative bacteria in the intestine of fish such as *Listeria*, *Bacillus*, and *Staphylococcus*. Gram-negative bacteria included certain species like *Aeromonas hydrophila*, *Citrobacter freundii*, *Escherichia coli*, *Enterobacter aerogenes*, *Vibrio anguillarum*, *Klebsiella* and *Pseudomonas* (Ngoc *et al.*, 2022) [19]. The presence of these bacteria in large concentrated levels in the liver, kidney, intestine, and other organs of fish could be a significant source of cross-contamination during processing procedure (Woraprayote *et al.*, 2018) [20].

Table (2) showed that the *coli* was only resistant to ampicillin in *Pangasius hypophthalmus*. These results were in agreement with recent research which reported that TEM-type β -lactamases are the most common plasmid-mediated β -lactamases in gram-negative bacteria and the most common in ampicillin-resistant *E. coli* isolates from foods and animals (Gonçalves *et al.*, 2013; Van *et al.*, 2008) [14, 15]. According to a study, the majority of the *E. coli* isolates showed resistance to oxytetracycline (97%), lincomycin (98%) and oxacillin (99%). Resistance to ciprofloxacin (59%) was more common than that to ampicillin (51%) and chloramphenicol (73%) (Abo-Amer *et al.*, 2018). *E. coli* exhibited the highest resistance toward streptomycin, which is followed closely by cloxacillin, oxacillin and erythromycin as reported by (Ayandele *et al.*, 2020) [17]. Another investigation reported that *E. coli* isolates was at

high level of ampicillin resistance (63.6%), that is followed by ceftazidime (69.09%), nalidixic acid (78.1%), ciprofloxacin (45.5%–49.1%), kanamycin (36.4%–41.8%), chloramphenicol, cefotaxime, meropenem, tetracycline, and colistin (Ngoc *et al.*, 2022) [19]. In a previous study reported by Salako *et al.*, (2020), it was found that 61-69% of *E. coli* isolates from *Pangasius* freezing processes at two factories in the Mekong Delta of Vietnam showed resistance to ampicillin (43-47%), followed by cefotaxime (33-40%). It was mentioned that the isolates prevalence of multi-drug resistance was evident. The development of antibiotic resistance bacteria was observed discovered from samples collected from *Pangasius* fish during farming could be interpreted by the buildup of antibiotics in the water, surroundings and ponds. The negative coagulase a level 1 score is given to this parameter related to *Staphylococcus* strains as hospital studies revealed increasing both incidences and infection rate with the types of bacteria, many of which are being antibiotic resistant, and because of unknown state where these *Staphylococci* strains originated or what the risky these strains are (Noseda *et al.*, 2013) [21]. In general, starving *Pangasius* fish prior to killing them could be efficient in decreasing the cross-contamination of fish fillet with a harmful, spoilage-causing, and bacteria that are antibiotic-resistant during processing (Ngoc *et al.*, 2022) [19].

Table 3: Percentages of antibiotic resistance in bacterial isolated in Zuri

Bacterial type	Ampicillin	Oxacillin	Gentamicin	Erythromycin	Ciprofloxacin	Tetracycline	Cefoxitin
<i>E. coli</i>	0	40	0	40	100	0	0
Enterobacter aerogenes	71.5	57.1	85.7	85.7	70	57.1	50
Coagulase negative staphylococcus	25	50	0	100	100	0	50
<i>Citrobacter braaki</i>	75	25	75	33	0	25	50
<i>Leuconostoc citreum</i>	50	50	0	50	25	50	50

The Black, Azov, and Caspian Seas' native cultriventris fish is named in Russia as "Zuri fish" or "Kilka fish." The both two species of anchovy and big eye kilka, *C. engrauliformis* (Borodin, 1904) and *C. grimmi*, are considered as the subspecies of this fish, which is regarded as a member of the Clupeidae subfamily of fish (Kessler, 1877) [22]. The Caspian Sea coast is a place where the common Kilka, *C. cultriventris caspi* (Svetovidov, 1941), can live. In comparison to other years, the harvest of this species of fish is increased due to the changing of the catch (Soleimani *et al.*, 2016) [23]. It was discovered that percentages of resistance to antibiotic in *E.coli* isolated in Zuri was (100%) to Ciprofloxacin and showed the lower resistant to (Oxacillin and Erthromycin), whereas there was no reported resistance to (Ampicillin, Gentamicin, Tetracycline and

Cefoxitin). On the other hand, *Enterobacter aerogenes* was resistant to Gentamicin (85.7%) and Erthromycin (85.7%), Ampicillin (71.5%) and Ciprofloxacin (70%), but revealed low resistance to Oxacillin, Tetracycline and Cefoxitin. Coagulase negative staphylococcus was of high resistance to Erythromycin (100%) and Ciprofloxacin (100%) while there no resistance to Gentamicin and Tetracycline was reported and low resistance to Ampicillin (25%), Oxacillin (50%) and Cefoxitin (50%) was observed. *Citrobacter braaki* was noticed to be resistant to Ampicillin (75%) and Gentamicin (75%) with showed low resistance to Oxacillin, Erythromycin, Ciprofloxacin, tetracycline and Cefoxitin. *Leuconostoc citreum* was of low resistance to all antibiotics table 3. This report could be considered the first study on zuri fish in Iraq.

Table 4: Percentages of antibiotic resistance in bacterial isolated in fillet fish

Bacterial type	Ampicillin	Oxacillin	Gentamicin	Erythromycin	Ciprofloxacin	tetracycline	Cefoxitin
<i>E.coli</i>	50	50	0	25	0	0	0
<i>Enterobacter aerogenes</i>	50	60	72	50	25	30	40
Coagulase negative staphylococcus	40	50	100	50	50	0	50
<i>Citrobacter braaki</i>	50	30	50	50	25	50	40
<i>Leuconostoc citreum</i>	45	30	0	50	25	100	50

Table (4) showed the Percentages of antibiotic resistance in bacterial isolated from fillet fish. *E.coli* exhibited low resistance to (Ampicillin, Oxacillin and Erthromycin) without resistance to (Gentamicin, Ciprofloxacin, tetracycline and Cefoxitin). *Enterobacter aerogenes* revealed low resistance to all antibiotics. While Coagulase negative staphylococcus was found to be resistant to Gentamicin (100%) without resistance to tetracycline (0%) and was of lower resistance to Ampicillin (40%), Oxacillin (50%), Erthromycin (50%), Ciprofloxacin (50%) and Cefoxitin (50%). *Citrobacter braaki* showed lower resistance to all antibiotic, but *Leuconostoc citreum* was observed to be high resistant to tetracycline (100%) with no resistance to Gentamicin, and of lower resistance to other antibiotics.

Coliforms, which involve *E. coli*, are microorganisms that may infect food and sicken people since they affect the digestive system. This is inescapable without following the right sterilization methods. In their study, Ellis *et al.* (2020) [27] noticed that the majority (60%) of the MDR isolates of *E. coli* showed co-resistance to following antibiotics such as ampicillin, ciprofloxacin, tetracycline, and trimethoprim in addition to resistance to other drugs. The all MDR isolates exhibited ciprofloxacin resistance without nalidixic acid resistance was reported in any of them. Prior to the national action plan which was implemented into place in 2016, tetracycline and other antimicrobial products were usually used in Vietnam, which could represent the high levels of tetracycline resistance (Longsand Lua, 2017) [28].

4. Conclusions and Recommendations

In conclusion, the findings of this study showed that a high-quality supply of animal protein may not be healthy and fit for human consumption because of sanitation lack, filthy equipment, and unhygienic cooking practices. Various species of bacteria isolates were found in samples collected from certain species of fish. These isolated bacteria showed different levels of resistance to various antibiotics. Good hygiene standards should be followed and recommended at the time of manufacturing fish, freezing, cooking. Furthermore, conservation procedures should be devised to

decrease food contamination by bacteria.

Conflict of Interest

Not available

Financial Support

Not available

5. References

- Balami S, Sharma, A, Karn R. Significance of nutritional value of fish for human health. Malaysian Journal of Halal Research. 2019;2(2):32-34.
- Shikha FH. 8. Post Mortem Changes in Snake Headed Fishes (*Taki*, *Channa punctatus* and *Shol*, *Channa striatus*) During Storage at Room Temperature (28 to 30° C) and in Ice. Journal of Agriculture, Food and Environment (JAFE) ISSN (Online Version): 2708-5694. 2022;3(1):45-53.
- Gelman A, Glatman L, Drabkin V, Harpaz S. Effects of storage temperature and preservative treatment on shelf life of the pond-raised freshwater fish, silver perch (*Bidyanus bidyanus*). Journal of food protection. 2001;64(10):1584-1591.
- Anagnostopoulos DA, Parlapani FF, Boziari zIS. The evolution of knowledge on seafood spoilage microbiota from the 20th to the 21st century: Have we finished or just begun?. Trends in Food Science & Technology. 2022;120(1):236-247
- Begum MD, Muniruzzaman M, Salauddin M, Rahman MM. Detection and antibiogram study of bacteria isolated from dried and cooked fish. Veterinary Sciences: Research and Reviews. 2021;7(2):134-142.
- FAO. Food and Agriculture Organization of United Nation. The State of World Fisheries and Aquaculture 2016 (SOFIA); c2016.p.176-182.
- Boylan S. Zoonoses associated with fish. The veterinary clinics of North America. Exot. Anim. Pract. 2011;14:427-438.
- Zaccone G, Capillo G, Fernandes JMO, Kiron V, Lauriano ER, Alesci A, *et al.* Expression of the

- Antimicrobial Peptide Piscidin 1 and Neuropeptides in Fish Gill and Skin: A Potential Participation in Neuro-Immune Interaction. *Marine Drugs*. 2022;20(2):145.
9. Novotny L, Dvorska L, Lorencova A, Beran V, Pavlik I. Fish: a potential source of bacterial pathogens for human beings. A review. *Veterinarni Medicina-UZPI (Czech Republic)*. 2004. Present status and future direction. *Aquac. Fish*. 2017 May 23.
 10. Yemmen C, Gargouri M. Potential hazards associated with the consumption of Scombridae fish: Infection and toxicity from raw material and processing. *Journal of Applied Microbiology*. 2022;132(6):4077-4096.
 11. Lehel J, Yaucat-Guendi R, Darnay L, Palotás P, Laczay P. Possible food safety hazards of ready-to-eat raw fish containing product (Sushi, sashimi). *Critical Reviews in Food Science and Nutrition*. 2021;61(5):867-888.
 12. Association of Official Agricultural Chemists, & Horwitz, W. Official methods of analysis. Washington, DC: Association of Official Analytical Chemists; c1975, 222.
 13. Handan-Dincer A, Baysal T. Decontamination techniques of pathogen bacteria in meat and poultry. *Crit Rev Microbiol*. 2004;30:197.
 14. Gonçalves A, Igrejas G, Radhouani H, Santos T, Monteiro R, Pacheco R, *et al.* Detection of antibiotic resistant enterococci and *Escherichia coli* in free range Iberian Lynx (*Lynx pardinus*). *Science of the total environment*. 2013;456:115-119.
 15. Van TTH, Chin J, Chapman T, Tran LT, Coloe PJ. Safety of raw meat and shellfish in Vietnam: an analysis of *Escherichia coli* isolations for antibiotic resistance and virulence genes. *International journal of food microbiology*. 2008;124(3):217-223.
 16. Abo-Amer AE, Shobrak MY, Altalhi AD. Isolation and antimicrobial resistance of *Escherichia coli* isolated from farm chickens in Taif, Saudi Arabia. *Journal of global antimicrobial resistance*. 2018;15:65-68.
 17. Ayandele AA, Oladipo EK, Oyebisi O, Kaka MO. Prevalence of multi-antibiotic resistant *Escherichia coli* and *Klebsiella* species obtained from a tertiary medical institution in Oyo State, Nigeria. *Qatar medical journal*. 2020;1:9.
 18. Salako DA, Trang PN, Ha NC, Miyamoto T, Ngoc TT A. Prevalence of antibiotic resistance *Escherichia coli* isolated from pangasius catfish (*Pangasius hypophthalmus*) fillet during freezing process at two factories in Mekong Delta Vietnam. *Food Research*. 2020;4(5):1785-1793.
 19. Ngoc TTA, Tu NC, Trang PN, Ha NC, Miyamoto T. *Pangasius hypophthalmus* Viscera as a Potential Vector of Bacterial Cross-Contamination and Resistance of *Escherichia coli* to Antibiotics. *Current Research in Nutrition and Food Science Journal*. 2022;10(2):711-719.
 20. Woraprayote W, Pumpuang L, Tosukhowong A, Zendo T, Sonomoto K, Benjakul S, *et al.* Antimicrobial biodegradable food packaging impregnated with Bacteriocin 7293 for control of pathogenic bacteria in pangasius fish fillets. *LWT*. 2018;89427-433.
 21. Nosedá B, Tong Thi AN, Rosseel L, Devlieghere F, Jacxsens L. Dynamics of microbiological quality and safety of Vietnamese *Pangasianodon hypophthalmus* during processing. *Aquaculture International*. 2013;21(3):709-727.
 22. Khalili H, Shabanipour N, Pournajafizadeh F. Structure and arrangement of photoreceptors in the retina of big eye kilka, *Clupeonella grimmeri* (Kessler 1877); c2014.
 23. Soleimani MR, Nikkhah M. Evaluation of antioxidant activity of protein hydrolysate from common kilka (*Clupeonella cultriventris caspia*). *Fisheries Science and Technology*. 2016;5(3):95-108.
 24. Hussein JA, Almosoy MT, Talal KA. The effect of supply and marketing on the chemical and bacteriological composition of frozen and marketed fish in the city of Baghdad, *Iraqi Journal of Market Research and Consumer Protection*; c2009.
 25. Falcao JP, Dias AMG, Correa EF, Falcao DP. Microbiological quality of ice used to refrigerate foods. *Food Microbiology*. 2002;19(4):269-276.
 26. Hameed Albassam N, Al-Doorri MM. November). Detection of Bacterial Contamination of Some Traditional Frozen Fish in Some Local Markets in Tikrit City. In *IOP Conference Series: Earth and Environmental Science*. 2021;910(1):012104. IOP Publishing.
 27. Ellis-Iversen J, Seyfarth AM, Korsgaard H, Bortolaia V, Munck N, Dalsgaard A. Antimicrobial resistant *E. coli* and enterococci in pangasius fillets and prawns in Danish retail imported from Asia. *Food Control*. 2020;14:106958.
 28. Long and Lua. Antimicrobial usage and antimicrobial resistance in Vietnam. Presentation; c2017.
 29. Mhango M, Mpuchane SF, Mpuchane BA. Incidence of indicator organisms, opportunistic and pathogenic bacteria in fish. *African Journal of Food, Agriculture, Nutrition and Development*. 2010;10:10.

How to Cite This Article

GM Baydaa, FA Jassim, KM Ibtisam. Identification and antibiogram of aerobic and anaerobic bacteria isolated from frozen fish retailed at public markets in Misan city, Iraq. *Journal of Advances in Microbiology Research*. 2022;5(4):01-03.

Creative Commons (CC) License

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 4.0 International (CC BY-NC-SA 4.0) License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.