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## Prevalence and distribution of etiological agent from open wound drainage

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#### Abstract

Wound infection due to various pathogenic microorganisms and the development of resistance to antibiotics is one of the major problems in the medical sector. This study aimed to identify the etiological agents of wound infection along with their antibiotic susceptibility. A total of 94 wound etiological agents were isolated from patients at microbiological laboratories for six months and analysed for the prevalence of bacterial as well as fungal isolates. The data was then compared with the survey data of the previous six months' bacteriological analysis reports.

Seven different bacterial strains were isolated from the wounds and also seven bacterial and 1 fungal strain were mentioned in the survey report. The prevalence of the microorganisms was then compared.

Keywords: Microorganisms, bacteria, prevalence, wound, resistance

### Introduction

The human body's primary organ and first line of defense, the skin covers the exterior of the body. Illnesses related to skin account for 34% of occupational problems worldwide. Skin diseases are the most common sort of disorder that afflicts people of all ages (Koshta and Sharma, 2023)<sup>[8]</sup>. The microbial pathogens triggering problems related to infected inflammatory diseases, human skin and soft tissue infections (SSTIs) after or while undergoing trauma, burn injuries, and surgical procedures forming pus or purulent wound drainage are of major concern. Wound drainage can be characterized as white to yellow fluid composed of necrotic tissues, cellular debris, and dead white blood cells (Hatlen and Miller, 2021; Ziesmer, 2023) <sup>[1, 2]</sup>. The etiological agents will remain commensal till the skin remains intact. Any abrasion on the skin surface provides an open door for bacterial invasion leading to infection (Maharjan and Mahawal, 2020)<sup>[9]</sup>. Both anaerobic and aerobic microbes have been associated with wound infections, which are frequent in healthcare facilities and can lead to serious morbidity, prolonged hospital stays, and infections that are resistant to multiple drugs (Puca et al., 2021)<sup>[3]</sup>. Furthermore, drains alone may not be effective, leading to discomfort and an extended hospital stay (Muthu et al., 2020)<sup>[4]</sup>. The rapid emergence of antibiotic resistance among potentially hazardous bacterial isolates and its appearance are regarded as serious dangers to public health globally (Chinemerem Nwobodo et al., 2022) [5]. There are a plethora of infectious bacteria as well as fungi, which can not only cause purulent infections but make them incurable hence creating major problems for patients as well as for clinicians (Dhara et al., 2023) [7]. Due to widespread misprescription and insufficient antibiotic dosage regimens, multidrug-resistant Gram-negative bacterial strains such as extended-spectrum beta-lactamases producing Enterobacteriaeae, Enterococcus spp., Klebsiella pneumoniae, Acinetobacter baumannii, E. coli, Pseudomonas aeruginosa and Gram-positive methicillin-resistant Staphylococcus aureus (MRSA) have been linked to pus infections in hospital settings across the past few decades (Prasanth et al., 2022)<sup>[6]</sup>. The objective of this study is to characterize the pyogenic bacteria from pus samples and to determine prevalence among all other species.

## **Materials and Methodology**

## **Research design**

We conducted a retrospective survey from records for microbial culture results reported in regional laboratories as well as isolated and identified the prevalence of the microorganisms from the purulent wound drainage (Shimekaw et al., 2022; Stout et al., 2023). No inclusion and exclusion criteria were put in the application.

### **Bacterial identification and characterization**

Specimens from regional laboratories were transported to the microbiology department laboratory at our University campus, for analysis within 2 h of collection in sterile swab tubes filled with 1 ml of brain heart infusion broth (Duzyol et al., 2020) [2]. Standard microbiological methods of cultivation have been employed to isolate and characterize fungi and bacteria. The media has been prepared following Cheesbrough guidelines. About 0.1 mL of purulent wound drainage from the open wound site of patients, respectively were inoculated into Brain heart infusion broth medium and incubated for 24 h (Bobai et al., 2022) [14]. Positive cultures were gram-stained and sub-cultured onto MacConkev agar plates or Mannitol salt agar plates respectively (Karah et al., 2020) <sup>[15]</sup>. Isolates were identified and processed according to standard techniques by Vitek 2 analysis (Al-Gburi and Mohammed, 2020)<sup>[16]</sup>. Data was interpreted according to the CLSI guidelines. Major bacterial isolates were evaluated for their percentage prevalence and compared with prevalence data obtained from the survey analysis.

### Data analysis

Data was analysed using Excel (Microsoft office, USA). Graphs or pie chart were used to show the prevalence and distribution of the isolated bacterial and fungal etiological agents against specimens (Pus, ear swab, sputum, stool, abscess, throat swab).

## **Results and Discussion**

The results of the survey analysis are represented here in Figure 1. The inner pie chart indicates the percentage of the pus sample reported in the six months survey of the percentage of the various samples reported in the survey during the phase of 6 months, the percentage of the pus sample reported was 68%, which is higher than any other clinical samples. In support of our results, Barai et al., 2021 <sup>[18]</sup> also reported 30.2% wound/pus swab from the samples collected between the 2001 to 2021. Whereas study of Jangla and Naidu (2018) <sup>[17]</sup> reported only 5.5% of wound swab and pus samples. The results of the prevalence analysis among the bacterial and fungal isolates indicates the incredible prevalence of the Staphyloccus auerus (306), followed by the Escherichia coli (137), Pseudomonas aeruginosa (119), Klebsiella spp.(62), Candida spp. (27), Streptococci spp. (20), Coagulase negative Staphylococcus aureus (12), Methicillin resistant Staphylococcus aureus (6), Aspergillus niger (1), (Figure 1). Whereas, Out of all 94 isolated bacterial strain, we observed the 57 Staphylococcus aureus, 12 Klebsiella pneumoniae, 11 Escherichia coli, 11 Pseudomonas aeruginosa, 1 Enterococcus faecalis, 1 Strenotrophomonas maltophilia, 1 Burkholderia spp., which again show the prevalence of the Staphylococcus spp. But have a distinct diversity among the etiology. Figure 2 is representing the percentage prevalence obtained from the experimental method.



Fig 1: Prevalence analysis of etiological agents from pus by survey method

The study of Biswas *et al.*, 2021 <sup>[19]</sup> on prevalence of methicillin resistant *Staphylococcus aureus* in pus samples in a tertiary care hospital of eastern India reported the 4.6% to 54.4% prevalence of methicillin resistant *Staphylococcus* 

*aureus.* Quayum *et al.*, 2022 <sup>[20]</sup> reported the 93.4% proportion of *Staphylococcus aureus* followed by the *Klebsiella pneumoniae*.



Fig 2: Prevalence analysis of etiological agents from pus by experimental method

Prevalence of multidrug-resistant (MDR) among the gramnegative bacteria causing pyogenic infections from June 2022-January 2023 reported *Pseudomonas aeruginosa* (42.9% cases), *Escherichia coli* (25.7%), and *Enterobacter* (7.1%) as the prevalent isolate, which is irreconcilable to our findings (Devi *et al.*, 2023) <sup>[21]</sup>.

## Conclusion

Results of the present study showed the significant prevalence of microbial strains within the pyogenic organisms. Particularly prominent among them is *Staphylococcus aureus* isolates responsible for purulent wound drainage and soft skin and tissue infections. Findings of the present study will be helpful for the clinician and researchers for identifying the highly prevalent strain, responsible for frequent incurable infectious wound drainage. Specific drug can be used to target the particular strains to cure such infections. This also highlights the urgency of specialized antibiotic to treat such persistent infections before it reaches the chronic stage.

## **Conflict of Interest**

The authors declare that no conflict of interests exist

## Reference

- Hatlen TJ, Miller LG. Staphylococcal skin and soft tissue infections. Infectious Disease Clinics. 2021 Mar 1;35(1):81-105.
- 2. Ziesmer J. Hybrid antibacterial microneedle patches against skin infections, doctoral dissertation, Karolinska Institutet (Sweden); c2023.
- 3. Puca V, Marulli RZ, Grande R, Vitale I, Niro A, Molinaro G, *et al.* Microbial species isolated from infected wounds and antimicrobial resistance analysis: Data emerging from a three-years retrospective study. Antibiotics. 2021 Sep 24;10(10):1162.
- 4. Muthu S, Ramakrishnan E, Natarajan KK, Chellamuthu G. Risk-benefit analysis of wound drain usage in spine surgery: A systematic review and meta-analysis with

evidence summary. European Spine Journal. 2020 Sep;29:2111-28.

- Nwobodo CD, Ugwu MC, Anie OC, Al-Ouqaili MT, Ikem CJ, Chigozie VU, *et al.* Antibiotic resistance: The challenges and some emerging strategies for tackling a global menace. Journal of Clinical Laboratory Analysis. 2022 Sep;36(9):e24655.
- Prasanth BK, Mahalakshmi K, Ramachandran U, Gurunathan N. Antimicrobial susceptibility pattern for managing maxillofacial Odontogenic infections-a tertiary care hospital-based study in Kanchipuram District Tamil Nadu. Neuroquantology. 2022;20(6):2486.
- 7. Dhara AK, Nayak AK, Chattopadhyay D, Editors. Antibiotics-Therapeutic Spectrum and Limitations. Elsevier; c2023 Jul 13.
- Koshta A, Sharma A. A Herbal Approach towards Skin Diseases: An Updated Review. International Journal of Newgen Research in Pharmacy and Healthcare. 2023 Jun 30:93-100.
- Maharjan N, Mahawal BS. Bacteriological profile of wound infection and antibiotic susceptibility pattern of various isolates in a tertiary care center. Journal of Lumbini Medical College. 2020 Oct 29;8(2):218-24.
- 10. Shimekaw M, Tigabu A, Tessema B. Bacterial profile, antimicrobial susceptibility pattern, and associated risk factors among patients with wound infections at Debre Markos Referral Hospital, Northwest, Ethiopia. The International Journal of Lower Extremity Wounds. 2022 Jun;21(2):182-92.
- 11. Stout L, Stephens M, Hashmi F. purulent skin and soft tissue infections, challenging the practice of incision and drainage: A Scoping Review. Nursing Research and Practice. 2023 Oct 6;2023.
- 12. Duzyol E, Aydogdu S, Duzyol M, Akgul N. Antimicrobial activity of root canal sealers on facultative gram-positive cocci. Int J Dent Sci Res. 2020 Apr 27;8:80-2.
- 13. Cheesbrough M. District laboratory practice in tropical

countries, Part 2. Cambridge University Press; c2005.

- 14. Bobai MA, Lawal DA, Nura SM, Joshua IA. Phenotypic and molecular characterization of Pseudomonas aeruginosa and Staphylococcus aureus isolated from patients' wounds in Barau Dikko Teaching Hospital, Kaduna, Nigeria. AJOPRED. 2022;14:096-118.
- 15. Karah N, Rafei R, Elamin W, Ghazy A, Abbara A, Hamze M, *et al.* Guideline for urine culture and biochemical identification of bacterial urinary pathogens in low-resource settings. Diagnostics. 2020 Oct 16;10(10):832.
- Al-Gburi A, Mohammed N. Isolation and molecular identification and antimicrobial susceptibility of Providencia spp. from raw cow's milk in Baghdad, Iraq. Veterinary Medicine International. 2020 Nov 19;2020.
- 17. Jangla SM, Naidu R. Study of Bacteriological profile and antibiotic sensitivity pattern in samples received from patients attending tertiary care hospital in Mumbai. Journal of Evolution of Medical and Dental Sciences. 2018 Jan 15;7(3):284-91.
- 18. Barai L, Saha MR, Rahman T, Sukanya M, Jafrin J, Ferdous J, *et al.* Trends in laboratory detection of Burkholderia pseudo mallei and their antibiotic susceptibility pattern from clinical samples over twenty one years from 2001 to 2021 in a tertiary care hospital of Bangladesh. Bangladesh journal of medical microbiology. 2021;15(1):8-14.
- 19. Biswas T, Chattopadhyay S, Mondal R. Prevalence of Methicillin Resistant Staphylococcus aureus in Pus samples in a tertiary care hospital of Eastern India; c2021.
- 20. Quayum A, Karmaker M, Ananna TY, Asna SM. Prevalence of MRSA, ESBL, and AMPC-betalactamase-producing bacterial profile in pus sample. 2022.
- Devi KD, Khandait M, Sharma M, Sardar M, Singh A, Saha R, *et al.* Prevalence of multidrug-resistant Gramnegative bacteria causing pyogenic infections at a tertiary care hospital in Haryana. MGM Journal of Medical Sciences. 2023 Jul 1;10(3):517-23.

#### How to Cite This Article

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